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# The first of five moults of *Forficula auricularia* L. (Dermaptera: Forficulidae)

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## **Abstract**

Most studies about the European earwig *Forficula auricularia* L. (Dermaptera: Forficulidae) assume that four larval moults occur from hatching to adult. In this note, we use observational and quantitative approaches to reveal the presence of an additional intermediate moult occurring immediately after egg hatching. We then briefly discuss how this finding may open up new possibilities for follow-up studies to improve our general understanding of the taxonomy, biology, and evolution of this species.

All insects pass through successive moulting events to reach adulthood. These moults typically allow general body growth and the development (or loss) of specific appendix and organs that become essential (or useless) at a certain period of their life (Chapman and Reynolds 2013). This is the case, for instance, of the genitalia and sexual ornaments that often appear only after the last moult into adulthood (Belles 2011; Rolff *et al.* 2019). The number and temporality of moulting events during insect development are under strong selection pressure, because they typically reflect age-specific physiological constraints and newly moulted individuals are often more vulnerable to pathogens and predation due to their resulting soft cuticle (Field *et al.* 1992; Corteel *et al.* 2009).

Somewhat surprisingly, the number of moulting events that occur during the development of the European earwig *Forficula auricularia* L. (Dermaptera: Forficulidae) remains unclear. This insect is present worldwide and exhibits a relatively long lifespan, with males and females living up to two years. Females typically produce a clutch of 30 to 60 eggs in early winter (Tourneur and Gingras 1992; Meunier *et al.* 2012; Tourneur and Meunier 2020), provide them with extensive forms of care through winter, and then remain with the newly hatched juveniles for several weeks during which they continue providing post-hatching care (Lamb 1976; Kölliker 2007; Koch and Meunier 2014; Diehl and Meunier 2018; Van Meyel *et al.* 2019). Whereas it is generally assumed that the number of larval stages present in this species is four (L1 to L4) and thus that larva pass through four moulting events during their lifetime (L1 to L2, L2 to L3, L3 to L4, and then L4 to adults; Jones 1917; Fulton 1924; Crumb *et al.* 1941; Lhoste 1942; Henson 1946, 1947; Behura 1956), the literature contains contrasting support for this claim. In particular, Chapman (1917a) mentioned the presence of six “ecdyses” while reporting only four moults in his subsequent paper (Chapman 1917b). Moreover, the occurrence of a moult immediately after egg hatching (a process commonly known as the intermediate moult; Chapman and Reynolds 2013) has been reported in a few other dermapteran species (Albouy and Caussanel 1990) such as *Marava arachidis* (Yersin) (Dermaptera: Spongiphoridae), where the first developmental stage only lasts the time of hatching (Herter 1943). Finally, a few studies report that *F. auricularia* larva are already well developed at the end of egg development (Fulton 1924; Behura 1956), suggesting that an intermediate moult could be possible just before or immediately after egg

hatching (Chapman 2013). Whether this moult indeed occurs in *F. auricularia* remained unknown (Lhoste 1942).

In this study, we used observational and quantitative approaches to demonstrate that an intermediate moult occurs immediately after egg hatching in *F. auricularia*. Our observations and measurements were based on 33 *F. auricularia* families originating from earwig females field sampled in August 2019 in Ripon, Québec, Canada (75° 14' 77" W, 45° 81' 06" N, altitude 195 m) and then maintained under standard conditions (Meunier *et al.* 2012). Of the sister species A and B currently included under the species name *F. auricularia* (Wirth *et al.* 1998), the population studied here belongs to subspecies A (Tourneur 2018). From field sampling to the day of oviposition, these females were fed with fresh carrots (Apiaceae) and an artificial diet (dry power of egg yolk 1/3, bee-collected pollen 1/3, and cricket 1/3) weekly. At oviposition, each female was isolated in a new Petri dish (diameter 10 cm) with a thin layer of humid sand. At that time, males and food sources were also removed to prevent egg cannibalism by males and because females stop foraging activity during the period of egg care (Kölliker 2007). After hatching, the larvae were maintained under the same standard conditions to conduct morphological measurements until they reached adulthood (see below).

Our observations of the intermediate moult began at the end of egg development. At that time, the eyes, mandibles, and *ruptor ovi* of the larvae present in the eggs are already visible (Fig. 1). The *ruptor ovi* (also called *egg burster*) is a tooth-like projection that an insect embryo or larva typically uses to break the embryonic envelope and egg chorion when hatching (Cooper 1966; Cai and Hua 2009). In *F. auricularia*, the *ruptor ovi* is located on the larval frons (Fig. 1A). We observed that this first moult begins with the rupture of the frontal and coronal sutures and occurs immediately after egg hatching. During this phase, the larva swallows a large quantity of air, which greatly distends its abdomen and thus facilitates the exuviation. As the exuviation takes place, the *ruptor ovi* initially present at the front position (of the larval stage thereafter called L0) then slides between the next stage's mandibles to the base of its abdomen (Fig. 2A to F). We propose naming the first stage "L0", both because such a pharate first-stage larva is not always considered as a larval instar in insects (Chapman and Reynolds 2013) and to remain consistent with existing literature referring to L1 to L4 larva in the European earwig (*e.g.*, Jones 1917; Fulton

1924; Crumb *et al.* 1941; Lhoste 1942; Henson 1946, 1947; Behura 1956; Vogelweith *et al.* 2017). The fact that the exuvia remains stuck at the base of the chorion facilitates the exuviation of the antennae, legs, and abdominal cerci. Once the larva is released (the frontal *oviruptor* is no longer present), it regurgitates its abdominal air and thus allows its abdomen, the intersegmental membranes of which have been distended, to return to normal size and shape. The sclerotisation of the cephalic capsule of the resulting L1 larva then begins. Our description of this additional moult in the European earwig fits with the description of intermediate moults reported in other groups of insects, such as Orthoptera and Hemiptera (Chapman and Reynolds 2013).

To confirm that the larvae visible in the eggs just before hatching belong to a different developmental stage (L0) than the larvae resulting from egg hatching (L1), we measured and compared the interocular distance of larvae from each of the newly defined five developmental instars. These measurements were conducted to the 0.01 mm using a binocular scope (Nikon-SMZ-2T; Nikon Canada Inc., Mississauga, Ontario, Canada) on 18 to 51 larvae per instar (Fig. 3). The eyespots are the first cephalic structures observable in the first developmental stage, and their measurement is possible through the larvae's translucent chorion (Fig. 1A). Our results confirm that the interocular distances of the measured larvae differed amongst the five developmental stages (linear model;  $F_{154,4} = 623.6$ ,  $P < 0.0001$ ) with mean values ( $\pm$  SE) of  $0.389 \pm 0.005$  mm in the newly defined L0 larva (in the eggs),  $0.495 \pm 0.004$  mm in L1,  $0.563 \pm 0.006$  mm in L2,  $0.681 \pm 0.013$  mm in L3, and  $0.834 \pm 0.005$  mm in L4 (Fig. 3). Each interocular distance differed significantly from another, including between the interocular distances of L0 larvae measured through the eggshell and of L1 larva measured at egg hatching (pairwise comparisons based on the estimated marginal means of the linear model presented above, with  $P$ -values corrected for multiple testing using Tukey methods; all  $P < 0.0001$ ; Fig. 3). Finally, these interocular distances (Fig. 1) fitted a straight line across the five newly defined larval stages ( $R^2 = 0.92$ ;  $y \sim 10.81 * x + 26.56$ ;  $x$  being the larval stage), a result in line with predictions of Dyar's Law (Dyar 1890). This law, which predicts that the head width of a larva should follow a regular geometrical progression in its successive instars, was originally formulated to define the number of moults across insects (Dyar 1890; Shull *et al.* 2010) and therefore confirms the presence of an intermediate moult in *F. auricularia*.

Overall, our data reveal the presence of an intermediate moult in *F. auricularia* (and thus a total of five, not four, moults, as commonly stated in the literature), which occurs almost immediately after egg hatching. This finding may open new opportunities for follow-up studies to improve our general understanding of the taxonomy, biology, and evolution of this species. For example, number of moults is an important criterion in taxonomy (Chapman and Reynolds 2013), and these results call for future work to explore whether this new information could be used to discriminate amongst the sister species A and B currently included under the species name, *F. auricularia* (Wirth *et al.* 1998). Similarly, natural variation in environmental constraints might affect the success, timing, and duration of this intermediate moult, as they do for any moulting event (Chapman 2013); our findings thus suggest investigations into the possible impacts of these constraints and of their potential stability in the invasion success of the European earwig worldwide (Quarrell *et al.* 2018; Tourneur and Meunier 2020). Finally, maternal care towards the eggs is both extended and essential in this species (Boos *et al.* 2014; Koch and Meunier 2014; Van Meyel *et al.* 2019); our results prompt further tests to shed light on this behaviour's possible impact on the success of this first moulting event (*e.g.*, Fig. 1B reports the case of a failed intermediate moult resulting in the death of a sclerotinised L1 larva in the eggs).

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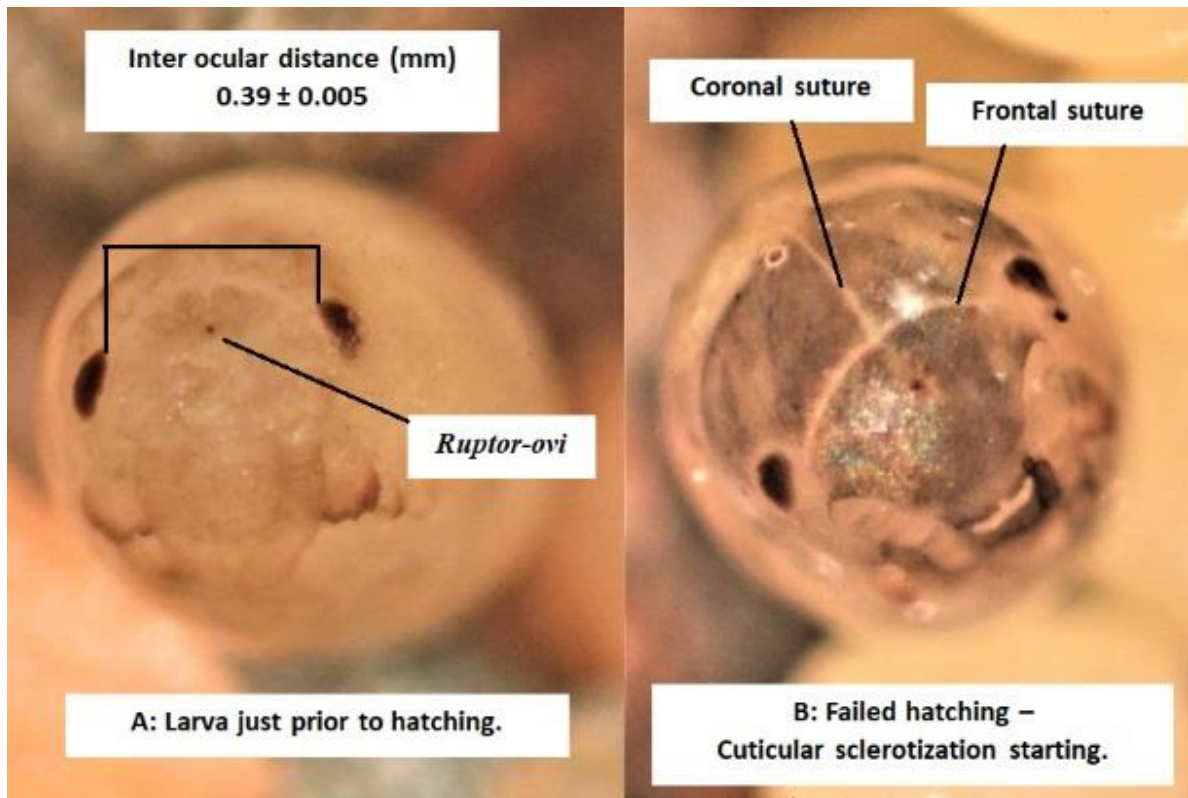
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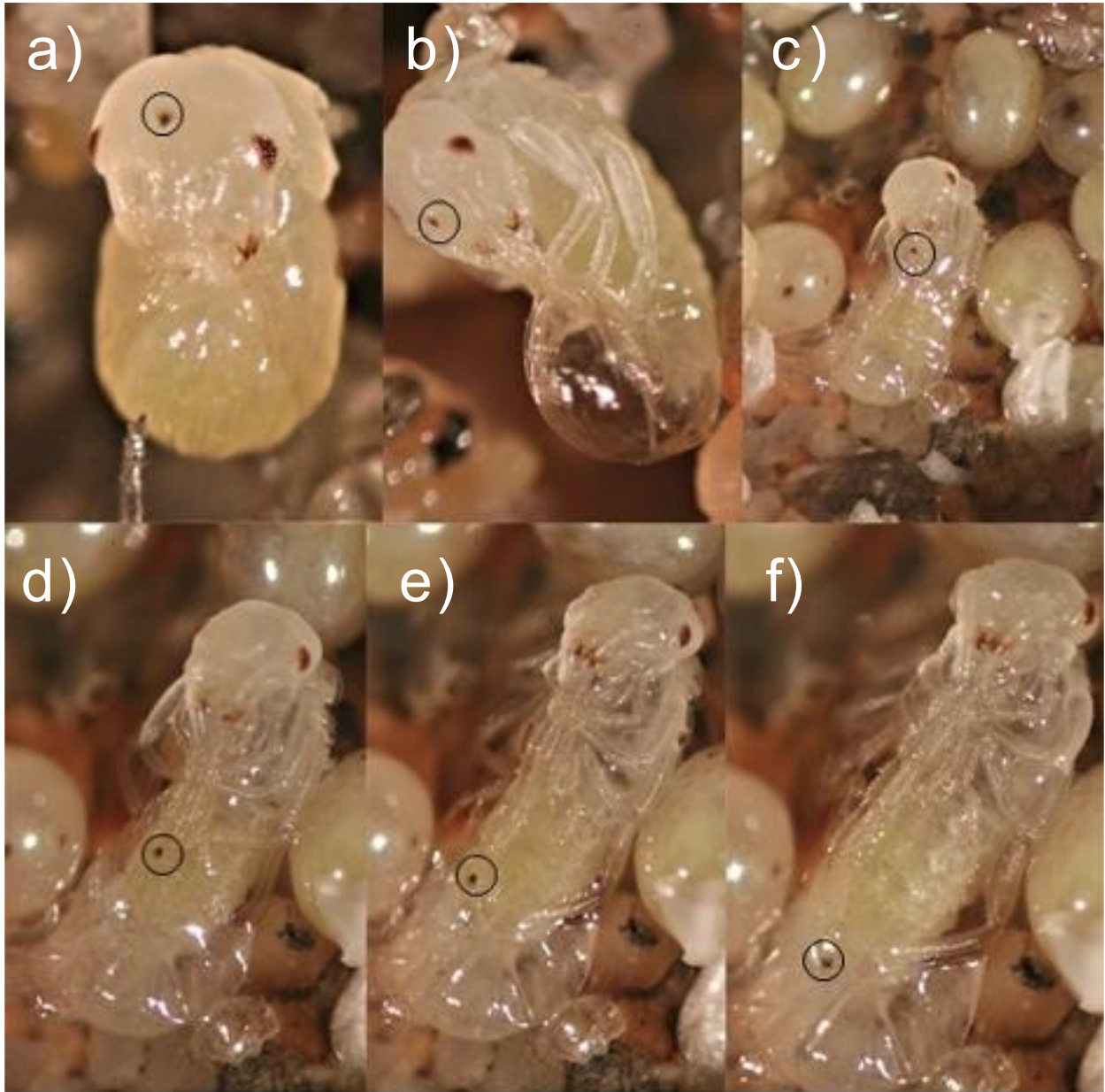
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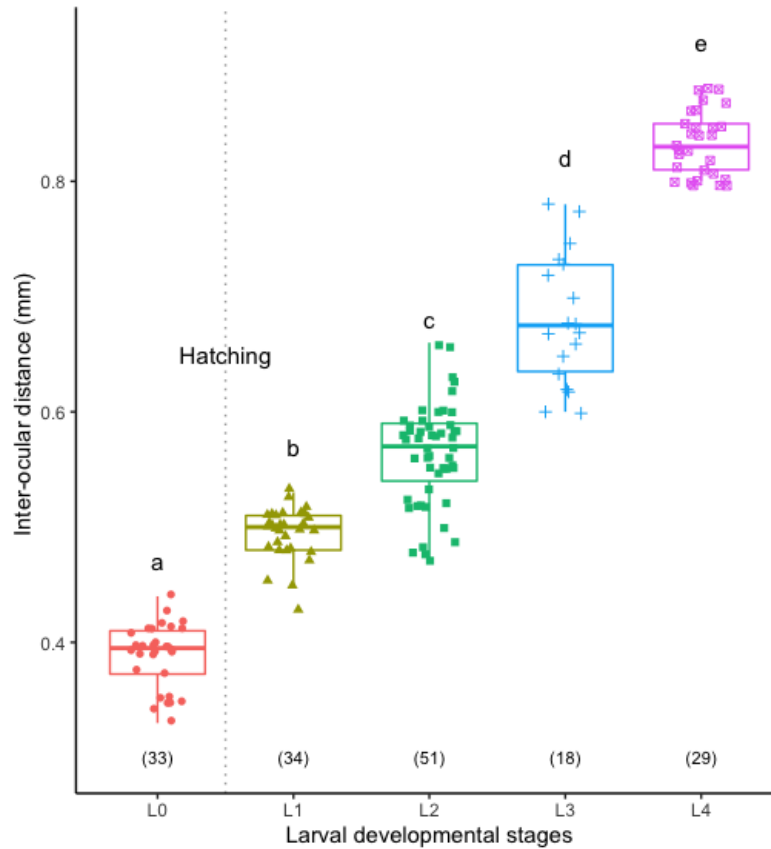
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**Fig. 1.** View of A) a L0 larva inside its egg; B) a dead L0 larva that did not manage to shed its embryonic cuticle and started cuticular sclerotinisation inside its egg.



**Fig. 2.** Migration of the *riptor ovi* after hatching and during first exuviation (from A to F). A circle locates the *riptor ovi* in each picture.



**Fig. 3.** Interocular distances measured in each of the five larval developmental stages. Boxplots depict median (bold bar) and interquartile range (light bar), with whiskers extending to 1.5 times the interquartile range and dots representing experimental values. Pairwise comparisons between developmental stages were conducted using estimated marginal means, and the P-values were corrected for multiple testing using Tukey methods. Different letters correspond to  $P < 0.0001$ . The number of measured larvae is indicated in parentheses at the bottom of each bar. The dashed line illustrates the period of egg hatching.